# **Course Specification**

University: Menoufiya Faculty: Science

Course Specifications:

Programme (s) on which the Course is given: MSC.cytology, Histology &

histochemistry

Major or Minor Element of Programmes: Major.

Department offering the Program: Zoology.

Department offering the Course: Zoology.

Qualifying course for M.SC. Students

Date of Specification Approval: 2013

#### **A- Basic Information**

Title: Embryo Culture Code: Z6416

Credit Hours: 2 Lecture: 1

Tutorial: 0 Practical: 2

Total: 3

## **B- Professional Information**

- **1- Overall Aims of the Course:** By the end of this course, the student should be able to:
- \* Recognize the different instruments and other requirements of embryo culture lab.
  - \* Stand upon the superiority of the holy creator.
  - \* Exploring different techniques used for embryo culture.

# 2- Intended Learning Outcomes of Course (ILOs):

# a Knowledge and Understanding:

# Student should acquire knowledge and understanding of:

- a1- The differences between Embryology and Embryo culture.
- a2- The related basic scientific principles and techniques.
- a3- The differences related to the culture of vertebrate embryos.
- a4- The related terminology, nomenclature and classification systems.

#### **b- Intellectual Skills:**

#### Student should be able to

- b1- Deduce the superiority of the holy creator.
  - b2- Differentiate between subject-related theories and assess their concepts and principles.
  - b3- Analyze, assess and interpret qualitatively and quantitatively science relevant data.
- b4- Identify the main aspects upon which vertebrate embryo culturing stands.
- b5- Construct several related integrated information to confirm, make evidence and test hypotheses.
  - b6- Think about mechanisms of embryonic development.

#### c- Professional and Practical Skills:

## **Student should be able to:**

- c1- Distinguish between different embryological courses.
- c2- Plan and report on the investigated data, using appropriate techniques and considering scientific guidance.
  - c3- Write notes on embryo culturing.
  - c4- Apply techniques and tools considering scientific ethics.
  - c5- Make clear labeled drawings for different developmental stages.

#### d- General and transferable Skills:

#### Student should be able to:

- d1- Monitoring different stages of embryo culturing.
- d2- Use effectively information and communication technology.
- d3- Development of scientific writing skills.
- d4- Acquire self- and long life-learning.
- d5- Think independently, set tasks and solve problems on scientific basis.
- d6- Work in group effectively; manage time, collaborate and communicate with others positively.
  - d7- Consider community linked ethics.

#### **3- Contents**

Торіс	No. of	Lecture	Tutorial /
	Hours		Practical
Instruments,			
Aseptic	6	2	2
techniques,	6		
Oxygen and			

carbon dioxide			
and life food.			
Culturing of			
amphibian larvae,	6	2	2
induced ovulation.			
Basic techniques			
for			
experimentation			
with amphibian			
embryos (Saline-			
removal of egg	6	2	2
membranes-			
narcosis-vital			
staining and			
grafting			
operation)			
Incubation of			
avian eggs,			
development in	6	2	2
the opened egg-	U	2	2
candling and			
making windows.			
Avian embryos	6	2	2
explantation	U		
Culturing of			
mammalian eggs			

and blastocysts-	6	2	2
Removal of the			
mammalian zona			
pellucida			
Normal tables of			
early embryonic	6	2	2.
development in	U	2	2
vertebrates			

# 4- Teaching and Learning methods

- 4.1- Lectures
- 4.2- Practical sessions.
- 4.3- Writing essays.
- 4.4- Oral presentation.

## 5- Student assessment methods

1- Essays to assess ability of writing

2- Oral exam to assess the degree of understanding

3- Mid-term to assess the degree of following up

4- Final exam to assess the whole performance.

## **Assessment schedule**

Assessment 1 Essay 1essay/term

Assessment 2 Oral exam Twice/ term

Assessment 3 midterm Week 7 Midterm

Assessment 4 final exam Week 15 final exam

# Weighting of assessments

## Weighting of assessments

Mid-term examination 20%

Final-term examination 40%

Oral examination 10%

Practical examination 20%

Semester work 10%

Other type of assessment 00%

Total 100%

## 5- List of references

- 5.1- Recent books in Embryo culture.
- 5.2- Scientific papers related to the subject of Embryo culture.
  - 5.3- Related web sites.

## 6- Facilities required for teaching and learning

- \* Lecture room provided with a white board.
- \* Dark room provided with a projector or data show.
- \* Student laboratory provided with oven, hot plate, incubator, microtome, and other laboratory facilities related to the subject of Embryo culture..

Course coordinator: Dr. Gamal M Badawy

Head of Department: Prof. Dr. Saber Sakr